

# Potential Applications for Using Stem Cells in Spine Surgery

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**Abstract:** While the use of biologics as adjuncts for spine surgery is growing annually stem cells have yet to be approved for this clinical application. Stem cells have the unique ability to differentiate into a variety of musculoskeletal tissues including bone or cartilage. Moreover they have been shown to secrete growth factors that promote matrix repair and regeneration and can down regulate inflammation and immune cell functions. It is these combined activities that make stem cells attractive candidates for advancing current techniques in spine surgery and possibly mitigating those pathologies responsible for tissue degeneration and failure thereby minimising the need for surgical intervention at a later date. This review focuses on the characteristics of progenitor cells from different sources and explores their potential as adjuncts for both current and future applications in spine surgery. Where possible we draw on the experimental outcomes from our own preclinical studies using adult mesenchymal progenitor stem cells, as well as related studies by others to support our contention that stem cell based therapies will play a significant role in spine surgery in the future.

**Keywords:** Stem Cell; Spine Surgery; Mesenchymal Stem Cell; Intervertebral Disc; Spine Fusion.

## INTRODUCTION

Spinal surgery is concerned with the bone-cartilage-neural interface. It is a field of surgery that is rapidly changing and evolving; not only with the development of novel techniques, approaches and devices but also with regularly emerging evidence from large clinical trials assessing its indications, efficacy and outcomes. The use of biologics in spine surgery has become widespread and, whilst stem cells are not yet in routine clinical use in spine surgery, it is likely that they will have a significant role in the future.

Stem cell science encompasses a wide range of different multipotential and progenitor cell types. Cells which may have potential application in spine surgery, and broadly classified as stem cells, have the ability to differentiate into tissues such as bone or cartilage and to secrete factors that promote matrix repair and regeneration. Furthermore, laboratory studies have shown that some such stem cells exhibit anti-inflammatory and/or immune modulatory properties. It is these combined characteristics that make stem cells prime candidates for advancing current techniques in spine surgery and for providing new strategies directed at targeting the underlying causes of spinal diseases and disorders.

This review will explore the characteristics of progenitor cells from different sources and focus on their application to both current and potentially future areas of spine surgery. In particular, different stem cell characteristics and results of their use in preclinical experiments will be discussed in relation to their potential for clinical translation in spine surgery. This review will not address spinal cord injury.

## STEM CELLS

Stem cells have two essential fundamental characteristics, the ability for self renewal, and the ability to differentiate into a variety of cell phenotypes [1]. Stem cells are loosely categorised as being either adult or embryonic depending on their origin.

## EMBRYONIC STEM CELLS

Human embryonic stem cell (ESC) lines were first derived from the blastocyst inner cell mass by Thomson [2]. Embryonic stem cells have a very strong capacity for self renewal and maintenance of viability in culture as well as the ability to differentiate into all three germ layer lineages; namely mesoderm, endoderm and ectoderm. Thus, they can differentiate into all cell types [3] but the former lineage is most relevant to spine surgery. Significantly, ESCs, by definition, have the potential to form teratomas [3] and there are moral and ethical dilemmas associated with their derivation from embryos [4]. We consider that ESCs will have a limited role in spine surgery in the foreseeable future, but may have a role in treating spinal cord injury [5].

## MESENCHYMAL STEM CELLS

Adult or somatic stem cells have been isolated from virtually all tissues in the body [6], however those most relevant to spine surgery are of mesenchymal origin. These cells do not share the same ethical dilemmas as ESCs as they are sourced from a range of tissues which themselves are not capable of embryogenesis. The bone marrow contains both mesenchymal stem cells (MSCs) and haemopoietic stem cells, the later have been used clinically for many years, predominantly in the treatment of haematological malignancy [7]. MSCs, as they are generally now called, are a clonogenic population of cells originally termed fibroblast colony forming unit (CFU-f) [8, 9]. Their discovery is often attributed to Friedenstein in the 1970s yet reports of the osteogenic potential of bone marrow cells can be found from the late nineteenth century [10].

Bone marrow is not the only source of MSCs, as they have also been isolated from a range of tissues including adipose tissue [11], synovium [12], muscle [13] and dental pulp [14]. Mesenchymal stem cells have a more limited ability for differentiation than ESCs [15]. Bone, cartilage, muscle and fat being the predominant end points of MSC differentiation [1], though recent reports have documented neural differentiation [16]. Mesenchymal stem cells have been defined by the International Society of Cellular Therapy by their characteristic plastic adherence, fibroblastic morphology and by cell marker expression of CD105, CD73, CD90 while lacking expression of CD45, CD34, CD14, CD11b, CD79a, CD19, HLA-DR [17].

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Reports comparing stem cells derived from bone marrow with those derived from adipose tissue have found differences in cell marker expression, such as CD49d and CD54 being present only in the latter [18]. Other studies have also documented large variability in marker expression of MSCs [19]. Cell surface marker expression also varies between passages of cells from the same isolate and a significant proportion of cells are quiescent [20]. The very nature of isolating cells by density gradient centrifugation and plastic adherence yields a heterogeneous population of cells even from isolates of the same source [21, 22]. It should be emphasised that the term MSCs refers to a loosely defined population of cells with variable characteristics [23]; the term MSC should be therefore be used with a degree of caution. Moreover there are differences in growth rates and differentiation potentials in cell populations harvested from different sources or by different methods [23, 24]. An example is that the growth factors required for a bone marrow derived MSC to differentiate into cartilage differ from the growth factors required for chondrogenic differentiation of an adipose derived MSC [25, 26]. It has also been shown that chondrogenic differentiation of adipose versus bone marrow derived MSCs occurs along different differentiation pathways [21] which is perhaps a reason why some believe adipose MSCs have an inferior chondrogenic potential compared with bone marrow derive MSCs [19, 27]. Thus, this issue of heterogeneity, in cell characterisation and differentiation, becomes significant if these cells are to be used clinically [28] in spine surgery, particularly in an allogeneic setting.

The fact that MSCs have been isolated from most tissues in the body begs an etymological question as to their origin and purpose. One hypothesis is that MSCs either originated from, or are synonymous with, endothelial progenitor cells lining blood vessels called pericytes and which have a vital role in tissue homeostasis and repair [25, 29]. However, the avascular nucleus pulposus of the intervertebral disc challenges the validity of this theory since it is devoid of blood vessels and thus pericytes, and yet it has been shown to contain so called, progenitor cells [30, 31]. These cells exhibit characteristics similar to MSCs, however small subsets were positive for the haematopoietic marker CD133. Interestingly, these CD133 positive cells were able to differentiate into bone and cartilage as were their CD133 negative counterparts. Intervertebral disc cells are also unique as they exhibit additional non-MSC characteristics such as the ability for phagocytosis [32], which suggests that the progenitors were laid down during ontogeny and are perhaps remnants from the embryonic origin of the intervertebral disc. Other progenitors cells which exhibit certain embryonic cell characteristics have also been isolated from adult body organs [33].

### MESENCHYMAL PROGENITOR CELLS

A population of stromal stem cells isolated from bone marrow [34] and adipose tissue [24] have been designated as Mesenchymal Progenitor Cells (MPCs). These are a purified monoclonal population of cells derived by immunoselection employing beads coupled to STRO-1 Mab (and other antibodies) to capture these precursor stem cells [35]. These MPCs contrast with the typical MSC populations derived using density gradient centrifugation followed by plastic adherence in culture which are a mixed cell population with limited potential for self renewal. The MPC are a richer source of CFU-F, not only with the potential for differentiation into the tissues of the mesenchymal lineage but also are more potent with respect to ability for self-renewal [34]. It is possible that those cells, which demonstrate differential potential within a typical MSC colony, are in fact MSCs.

### AMNION EPITHELIAL CELLS

Between the extremes of ESCs on one hand, and adult stem cells on the other are a diverse range of other pluripotent stem cells. These are mainly derived from pregnancy tissue such as the fetus [36], amniotic fluid [37], placenta [38], fetal membrane [39] and

umbilical cord [40]. There is great interest in cells from placenta and fetal membrane as, like the MSC and in contra-distinction to the ESC they do not incite ethical dilemmas; being derived from tissue that is usually discarded after birth. They share with ESCs the ability to differentiate into all three lineages but, unlike ESCs, do not form teratomas [39, 41]. There are also MSCs present in after-birth tissue such as in the chorion and Wharton's jelly [42, 43]. These MSCs, together with the other placental pluripotent cells, also have the advantage of being obtained without the need for an invasive procedure such as bone marrow biopsy required for harvesting bone marrow derived MSCs. Cells of these origins are already used clinically for treating a range of haematological conditions such as malignancy [44].

Our group has an interest in cells derived from the amniotic membrane obtained from term deliveries, which have been termed amnion epithelial cells (AECs). These cells are originally derived from epiblast cells prior to gastrulation which migrate along the walls of the amniotic cavity to form the amnion epithelium [39]. Thus they possess pluripotent characteristics similar to ESCs. Amnion epithelial cells can differentiate down all three lineages [41] and the ability to evade an immune response with allogeneic or xenogeneic transplantation due to minimal MHC class one and two expressivity [45]. Moreover, such cells have been shown to have anti-inflammatory properties [46]. Although, amnion has been used clinically for the treatment of burns and ocular injury [47-49], AECs have not been used clinically in spine surgery. Our group are currently studying their effectiveness in spinal fusion in preclinical trials [50].

### OTHER STEM CELLS

Historically, differentiation was thought to be an irreversible unidirectional process such that an MSC was the progeny of an embryonic cell that had differentiated and was now committed to the mesenchymal lineage. This edict, however, has now been discarded since *in vitro* studies have shown that MSCs in culture induced to differentiate into adipocytes, osteoblasts or chondrocytes can be trans-differentiated to an alternate cell type simply by changing the culture media and constituent growth factor conditions [51]. The demise of the unidirectional hypothesis was also confirmed by studies on induced pluripotent stem cells (iPS). These studies demonstrated that fully differentiated somatic cells, such as dermal fibroblasts, could be re-programmed into cells with morphological, antigenic and epigenetic characteristics of an embryonic stem cell with pluripotent potential [52]. Currently, however, there is limited clinical utility of this finding as, like ESCs, iPS cells have the ability to form teratomas and the potential for oncogenesis [33, 52-55]. An alternate approach may be using partially differentiated iPS cells. In a murine model of hindlimb ischaemia iPS-derived MSCs had superior effects compared with bone marrow derived MSCs and no tumour formation was evident at four months [56].

Other adult stem cells that have been recently described include marrow isolated adult multilineage inducible stem cells (MIAMI cells) [57], and multi-potent adult progenitor cells (MAPC) [58]. These cells are thought to be a precursor to bone marrow cells including the hematopoietic and mesenchymal stem cells and thus have a differentiative potential compared to classical MSCs [59, 60]. Another progenitor cell, termed Very Small Embryonic-Like Cells (VSEL) have been isolated from a range of tissues [33]. Of great interest is that VSEL cells have been found to be circulating in patients following significant biological insults such as myocardial infarction and stroke [33, 61]. To our knowledge none of these cell types have not been tested in spine surgery.

### CLINICAL APPLICATION OF STEM CELLS

Of the cells discussed thus far, we believe the MSC class of cells, and in particular MPCs, due to their well-characterised and

tested monoclonal population, are currently closest of all stem cell types to mainstream clinical use in augmenting spine surgery.

The mode of action by which a stem cell may augment spinal surgery is threefold. Firstly, as already discussed, they possess the ability to differentiate into bone and cartilage, which is an important attribute for fusion surgery and disc regenerative therapies respectively. Secondly, they secrete multiple bioactive factors such as multiple bone morphogenetic proteins (BMPs)<sup>1</sup>. Bone morphogenetic proteins have been used clinically to promote osteogenesis [62, 63] and preserve disc integrity [64]. Thirdly, MSCs possess immune modulatory, anti-inflammatory and anti fibrotic activities. The expression of these effects in-situ would have enormous potential in the treatment of both myelopathy and radiculopathy. This being said, it is highly likely that a multimodal approach may be adopted in the future where two or more cell types with different characteristics and potentials may be used in combination to achieve different effects. An example may be to promoting fusion using the osteogenic properties of cell A together with treating radiculopathy using the anti-inflammatory properties of cell B.

### MESENCHYMAL CELL TRANSPLANTATION

Spine surgery has been utilising autologous mesenchymal cell transplantation for many years. Iliac crest autograft is still considered the gold-standard source of bone graft by many as it is a) osteogenic (cells within the graft can directly differentiate into osteogenic cells), b) osteoinductive (factors and cells within the graft can signal endogenous local cells to differentiate into osteogenic cells) and c) osteoconductive (the graft itself can act as a scaffold for bony in-growth) [65]. The bone marrow stroma that is transplanted during these procedures is responsible for its osteoinductive properties presumably due to the presence, albeit in low numbers, of stromal progenitor cells. Despite this, alternatives to autograft have been sought to minimise potential donor site morbidity [66]. Autologous mesenchymal cell transplantation has a similar potential for donor site morbidity, but the bigger issue is that, following harvest, the cells require culture expansion. This introduces cost of individual culture expansion and logistical impediments, as surgery using these cells is delayed by several weeks from the time they are harvested. Furthermore autologous cells are of variable quality depending on the protoplasm of the patient.

Allogeneic cell transplantation overcomes these problems as an 'off the shelf' product with batch to batch consistency and can be used as needed. The potential for transmission of infection and rejection are the obvious concerns of an allogeneic approach. Donor screening and extensive testing of the cells, in a similar manner employed for blood transfusions minimises the infection risk. The rejection risk is minimal as mesenchymal cells are of low immunogenicity. They exhibit low levels of cell surface markers such as the MHC class and lack surface expression of immune co-stimulatory molecules [67]. MSCs also lack the ability to induce an allogeneic Mixed Lymphocyte Reaction (MLR) [68, 69] and secrete multiple anti-inflammatory and immunosuppressive cytokines, e.g. IL-10 [68]. They actively suppress ongoing immune reactions by modulating dendritic cells and preventing monocyte and macrophage differentiation and activation [69, 70]. Allogeneic transplantation of the MSC class of cells is therefore an attractive prospect for spine surgery, with the potential to revolutionise this field.

### SPINE SURGERY:

#### Current Indications

The indications for a spinal operation are invariably pain and/or disability for which conservative measures have been exhausted. Degenerative disease is the most common underlying aetiology but congenital and traumatic conditions also play a causative role.

The most common degenerative conditions are disc herniations and spondylosis, both of which occur predominantly in the cervical and lumbar spine [71]. Both of these conditions usually manifest clinically as they cause neural compression. This may be central compression, which affects the spinal cord in the cervical region or the cauda equina in the lumbar region and/or nerve root compression. Nerve root compression generates radicular pain and may be associated with other features including weakness and sensory disturbance, collectively termed radiculopathy. The term compression, which is often used to describe the cause of radiculopathy, implies a mechanical aetiology, however, it is well accepted that a chemical or inflammatory component plays an important role in nerve root irritation and hence the causation of radiculopathy [72, 73].

The pain caused by gradual compression of the cauda equina is termed neurogenic claudication. This is a dynamic phenomenon of pain on walking or standing that is relieved by flexion [74]. Compression of the spinal cord in the cervical region causes cervical myelopathy which, unlike in the lumbar region, may not produce pain at all [75]. Cervical myelopathy causes progressive neurological symptoms such as disturbance of gait or hand dexterity [76] which are the most devastating sequelae of degenerative disc disease [77].

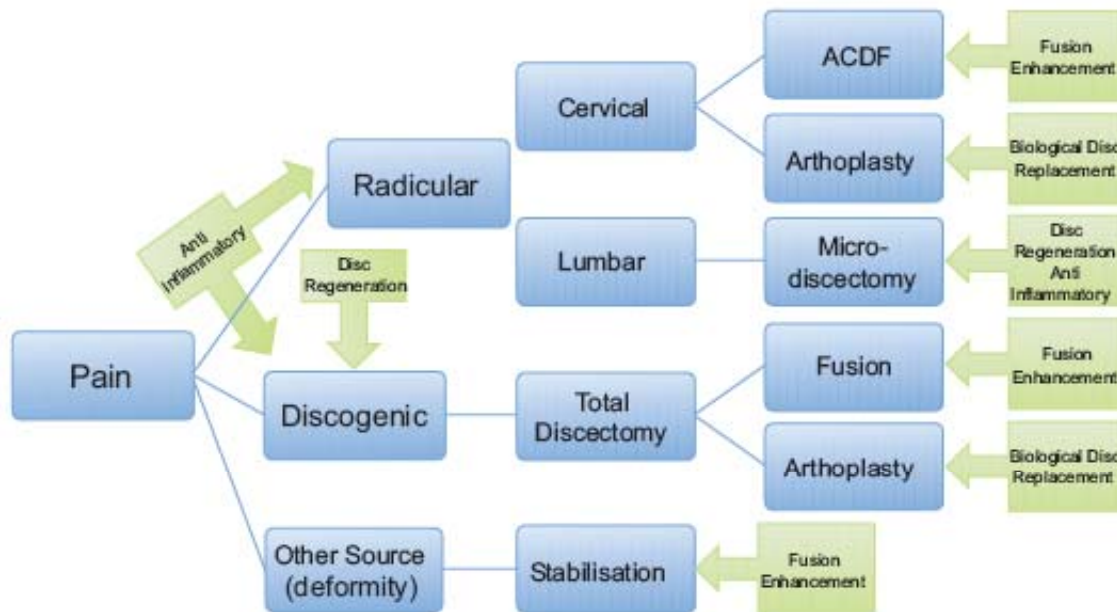
Pain frequently occurs without neural compression as the degenerate disc may itself be a pain generator causing what has been termed 'discogenic pain' [78, 79]. Discogenic pain is more difficult to diagnose than radicular pain as the imaging features of an ageing compared with a degenerate or painful disc are often indistinguishable [79-82]. Facet arthropathy, deformity and segmental instability arising from congenital or traumatic causes may also precipitate pain either themselves or through neural compression. Back pain may also arise from other extra axial sources arising from dysfunction in other joints such as the hip [83].

Surgery to the spine generally aims to fulfil one or more of the following goals: neural decompression, stabilisation or restoration of the deformity and removal of the pain generator. A summary of the current surgical approaches and where we believe stem cells may augment these approaches or obviate the need for surgery are shown in Fig. (1).

### LUMBAR VERSUS CERVICAL

The operative treatment of discopathies in the cervical and lumbar spine is significantly different and this impacts on the choice of stem cell to be used for each region. Whilst there is some controversy regarding the surgical approach [84] and the amount of disc to be removed [85], a subtotal discectomy *via* a posterior approach is the most commonly employed technique for a lumbar disc herniation [85, 86] Fig. (2), whereas the majority of spine surgeons would perform a total discectomy *via* an anterior approach for treatment of cervical radiculopathy and this is often followed by interbody fusion, hence anterior cervical discectomy and fusion (ACDF) [87-89] (Fig. 3). This distinction is largely due to anatomical differences and, although surgery always aims to minimise neural retraction, the spinal cord has much less tolerance to retraction in the cervical region compared with retraction of the nerve roots in the lumbar region; consequently the posterior approach in the lumbar region which invariably requires a degree of neural retraction. A subtotal lumbar discectomy in the current era is typically performed with operative magnification hence the term lumbar microdiscectomy, which will be discussed later [90]. Total discectomy and interbody fusion in the lumbar spine is reserved for patients with instability or deformity and most commonly for patients suffering chronic axial low back pain [91, 92] as the discectomy removes the pain generator [93]. This may be approached through an anterior lumbar interbody fusion (ALIF), posterior lumbar interbody fusion (PLIF), transforminal lumbar interbody fusion (TLIF) or extreme

<sup>1</sup> Zannettino personal communication



**Fig. (1).** Schema of current surgical indications and operations (blue boxes) and potential roles for stem cell based therapies (green boxes with arrows).

lateral interbody fusion (XLIF). There is strong evidence supporting surgery [92] for refractory back pain in carefully selected patients [91, 94].

### INTERBODY FUSION

Following total discectomy the void in the interbody space needs to be stabilised. The current options include prosthetic disc arthroplasty or fusion. In the cervical region there are some who advocate discectomy alone [95-99] which usually results in fusion anyway but has the disadvantage of reducing foraminal height [98]. Systematic literature reviews have failed to identify the best approach from published trials [87, 100], and therefore an ongoing prospective trial is underway that aims to answer this question [101].

Disc replacement with a new tissue engineered disc, using stem cells, is certainly the Holy Grail in terms of future therapies and would likely provide the best option for occupation of the interbody space following discectomy. This being said, it is likely that for certain indications, such as in severe traumatic injuries, such treatment may not be feasible and spinal fusion will therefore likely always remain part of the surgical armamentarium. Regenerative therapies using stem cells may also not be possible in patients who have severe disc degeneration, in whom the aetiological factors such as nutritional impairment to the disc persist, such as is the case with severe calcification of the endplates [102]. The use of stem cells in such an interbody space would be futile as this is an extremely hostile environment, and without adequate nutrition they would not survive [103]. Fusion therefore, may be the only option for this group of patients.

There are various graft options currently available for both cervical and lumbar fusion surgery. Tricortical autograft bone, usually from the iliac crest was the first interbody implant used, achieving high fusion rates, for the reasons discussed above. Significant problems with autograft can occur; including donor site residual pain, infection and cosmetic problems [66]. Allograft of cadaver bone is therefore used to avoid donor site morbidity although with an inferior fusion rate compared with autograft [104]. Allograft has potential problems of its own including rejection, resorption, infection and logistic issues [105]. Alternative synthetic interbody products such as interbody cages (Fig. 3) filled with bone substitutes such as

tricalcium phosphate are widely available producing successful fusion without donor site morbidity [106, 107]. These bone substitutes are osteoconductive *per se* meaning they provide a matrix into which local cells including endogenous mesenchymal stem cells, blood borne cells and osteoblasts can integrate and produce bone. These substitutes lack the osteoinductive ability that autografts have to a small extent due to the presence of bone marrow stromal cells in the graft.

There has long been a need for factors that could be used to increase fusion rates in spinal surgery. This is particularly so for lumbar surgery and for ALIF where fusion rates are lower. The situation is exacerbated in patients with comorbidities such as rheumatoid arthritis [108], smoking [109, 110] or patients on anti-inflammatory medications [111] which can independently decrease fusion rates. Recombinant bone morphogenetic protein-2 (BMP-2) and recombinant BMP-7 have been widely used in a range of spinal and other orthopaedic surgeries as osteoinductive agents to promote fusion [112-115]. There are, however, reports of adverse effects of their use in the cervical spine where ectopic bone formation and soft tissue swelling have been reported [116, 117]. Recently, the Food and Drug Administration (FDA) issued a warning advising that rhBMP should only be used in an approved clinical trial in the cervical spine [104, 118].

A recent preclinical study by our group designed to assess the safety and efficacy of MPC facilitated cervical interbody fusion [119] showed no cell related adverse events, including absence of swelling, airway compromise or neural compression [120]. MPCs were added to a commercially available tricalcium phosphate and hydroxyapatite carrier and were demonstrated to promote a faster and more robust fusion than current clinical treatments using autograft or carrier alone [120-122]. Biologically there is an interplay between the 14 naturally occurring BMPs involved in osteogenesis [123] and MPCs have been shown to secrete many of these growth factors<sup>2</sup>. This paracrine effect of MPCs, in addition to a direct effect of osteogenic differentiation at the fusion site, could account for the beneficial effects mediated by these cells in our animal model [120]. These observations support our contention that stem cells, such as MPCs, are likely to play an important role in fusion surgery

<sup>2</sup> Zannettino personal communication

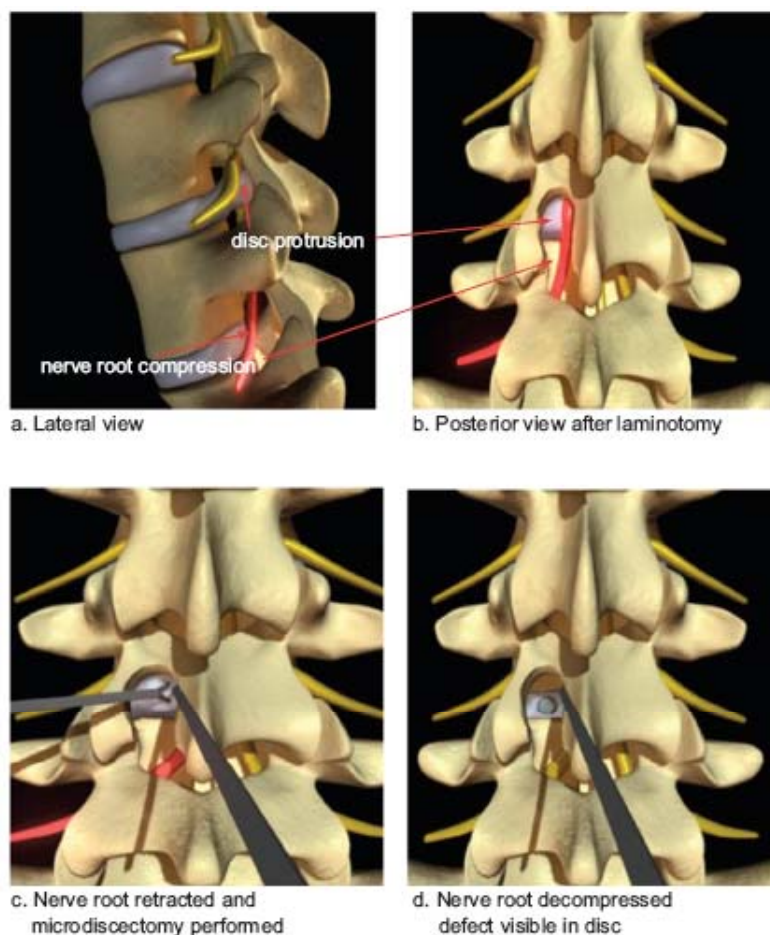


Fig. (2). Lumbar disc pathology.

in the future, particularly in the cervical spine where there is currently no satisfactory alternative. Clinical trials to assess the safety and efficacy of MPCs for cervical and lumbar interbody fusion are presently underway [124, 125].

### POSTEROLATERAL LUMBAR FUSION

A lumbar interbody fusion, when performed from the posterior or transforaminal approach, is invariably accompanied with posterolateral-instrumented fusion. This is a bony fusion in the posterolateral gutters, typically undertaken in conjunction with instrumentation such as a screw-rod construct inserted trans-pedicularly. A posterolateral fusion may also be performed without an inter-body fusion for trauma or deformity correction such as in cases of scoliosis or spondylolisthesis. Bone grafts, graft substitutes and osteoinductive agents are used widely posterolaterally in the lumbar spine [126, 127]. The graft volume required in posterolateral fusion is substantially larger than the volume required for interbody fusion and the distance for bony bridging to achieve fusion is also larger. Mixed results have been reported using BMPs in posterolateral fusion [63, 128, 129], however, the limited number of clinical trials undertaken to assess the efficacy of MSCs and MPCs for posterolateral lumbar fusion have shown promising outcomes to date [130, 131].

### MICRODISCECTOMY

Lumbar microdiscectomy involves partial discectomy or removal of the offending disc fragment from the peri-discal space [85, 132]. In this procedure the majority of the disc remains in place after surgery since at the onset of the disc herniation it is generally

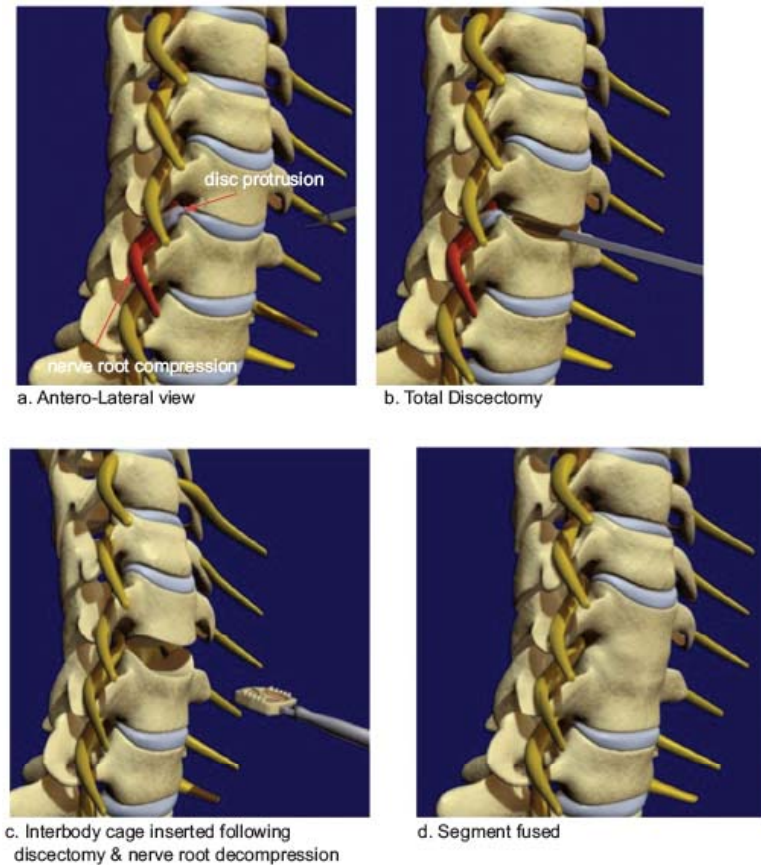
part of the nucleus pulposus and some annular fragments that are sequestered extradiscally [133]. Whilst microdiscectomy normally resolves the patient's pain in the short-term there is nevertheless a significant incidence of recurrent disc prolapse or discogenic back pain arising from the damaged disc structure, which does not undergo spontaneous healing [134-136].

The long-term problems associated with microdiscectomy, together with a more conservative approach to spinal surgery in recent years, has stimulated increased interest and research into methods that might be harnessed to promote the repair and regeneration of the injured disc. Mesenchymal stem cells have played and will continue to play a pivotal role in these endeavours.

Cells from the annulus fibrosis [137] (AF) and nucleus pulposus [138] (NP) have been isolated from explanted discs and cultured both *in vitro* [139, 140] and *in vivo* [141] within a variety of scaffolds. Stimulation in the form of growth factors or mechanical strain [142] have been tested for their ability to facilitate cell proliferation and even cells from severely degenerated discs showed some positive response in culture [143, 144]. Other experiments have resulted in differentiated disc like cells from MSCs [145] or from co-cultures of MSCs with disc cells [146]. These reports demonstrate that, even in the diseased state, AF and NP cells retain the ability to be activated and that regeneration of extracellular matrix constituents is possible. Moreover, MSCs have been co-cultured with whole disc tissue explants, rather than with isolated cells and this produces functional disc-like cells and extracellular matrix [147].

The Eurodisc study specifically investigated regenerating the disc following microdiscectomy in a clinical trial [148-150]. It





**Fig. (3).** Cervical disc pathology.

showed a decrease in back pain using autologous chondrocyte harvested during microdiscectomy and later transplanted following expansion. The limitation of this technique is the need for culture expansion of explanted cells and their injection into the damaged disc as an additional procedure. An alternative approach aimed at injection of regenerative stem cells during a microdiscectomy procedure would be beneficial and preferable to an approach requiring metaphyseal procedures. Preclinical studies using autologous adipose derived mesenchymal cells [151] or allogeneic MPCs [152] have demonstrated the regenerative potential of this approach but the former required an additional harvesting procedure pre-microdiscectomy highlighting the potential advantage of using allogeneic MPC for such therapies.

### DISC ARTHROPLASTY

Whilst the success rate of surgically relieving discogenic pain is high in the correctly selected patient, discectomy (or removal of the 'pain generator') and fusion of this segment may exacerbate degeneration at adjacent levels, a problem known as adjacent segment disease [153]. Motion preservation techniques or disc arthroplasty using a variety of implantable devices are already used in selected patients in an attempt to reduce the incidence of adjacent segment disease. Whether there is a long term benefit of disc arthroplasty presently remain unclear [101, 154-158]. Although lumbar disc arthroplasty has perhaps more supportive evidence than cervical, in general, longer term studies are needed. Nevertheless, disc arthroplasty prostheses are expensive and non-biological. The capacity to insert a new tissue engineered disc following discectomy, if this were possible, could provide a superior approach to the use of existing fusion or prosthetic procedures.

Despite all the recent progress in the use of tissue engineering in surgical spinal disc repair, the ability to tissue engineer a new

disc is still some way off, in particular, engineering a biomimetic annulus fibrosis presents the greatest challenge [77]. An alternate approach that has been used is allograft disc transplantation [159]. In one report cervical discs together with the endplates and uncoversal joints were removed from donors and frozen and then subsequently transplanted into patients. In this series of five patients with a five-year follow up, there was radiological evidence of fusion over the transplanted disc space in some cases [159]. As no explants of the transplanted discs were available, disc analysis could not be performed, but it is possible that a fibrous non-union was achieved over the interbody space [159]. The authors themselves conceded that there was no way of establishing whether the annulus and nucleus cells in the transplanted discs survived the transplantation [159]. Motion preservation however was maintained to some extent in all patients and there was no evidence of adjacent segment disease in the interim results reported.

The use of stem cells together with an appropriate chondrogenic stimulus and an appropriate bio-scaffold to replace a damaged or degenerated disc could be implanted to provide a fibrous or cartilaginous joint similar to that achieved in the transplantation study above. The tissue generated by such a procedure would not, of course, reproduce the complex integrated matrix of the disc but could provide a cartilaginous structure, which would provide articulation of the adjacent vertebral bodies to achieve similar results. Our group is currently exploring such an approach *in vivo* [160, 161].

### PAIN AND INFLAMMATION

Pain is the most common underlying reason and presenting feature for spine surgery and the goal of the above mentioned procedures is to alleviate pain through neural decompression or re-

removal of the pain generator [162]. Since inflammation is a causative factor of this pain [72, 73], the anti-inflammatory properties of MSCs have the potential to directly reduce pain in addition to their other applications in the surgical procedure. MSCs act through various anti-inflammatory mechanisms. They do not induce an allogeneic Mixed Lymphocyte Reaction (MLR) [45, 68, 69], they secrete multiple anti-inflammatory and immunosuppressive cytokines, such as interleukin-10 [68] and they actively suppress ongoing immune reactions by modulating dendritic cells and preventing monocyte and macrophage differentiation and activation [69, 70].

#### FUTURE TREATMENTS

The biologic approaches discussed in this review using stem cells, if successful, would provide alternative options and improvements to current techniques used in spinal surgery. Moreover, we consider that the use of stem cell to treat spinal disorders offer a far more exciting and diverse role than do growth factors which have short half-lives and can induce untoward effects in sites adjacent to their application. Stem cells offer the opportunity to treat the pathological defects at an earlier time point than is currently possible, halting or reversing further degeneration consequently obviating the need for surgery all together. In this respect, our own research has generated histological and biochemical evidence of tissue regeneration, with corollary imaging improvement, following injection of stem cells into degenerative lumbar discs in an ovine model [163]. Others too have provided *in vivo* evidence of disc regeneration using stem cells in other models [151].

These promising experimental studies in laboratory animals are still in the process of translation to human clinical trials. The major clinical challenge will be the ability to diagnose the 'pain generator' in the patient presenting with back pain. Notwithstanding the use of multiple modalities [164], including clinical examination, imaging (usually MRI) and discography [165, 166], distinguishing the ageing versus the degenerate disc is still difficult [80, 167]. The pain can arise from a source other than the disc [83] and, if it is from the disc, its causation may be from pathological in-growth of nerve fibres [78, 168], which is not something that was specifically tested in the disc regeneration animal models. The known anti-inflammatory/immunosuppressive effects of some stem cells are likely to have a positive effect in spinal application and although there is evidence in animals of pathological ingrowth of nerve fibres [163], the true analgesic effect of stem cell therapy can only be truly tested in human clinical trials.

#### CONCLUSION

In summary, stem cell science holds much promise in complementing, improving and augmenting current techniques in spinal surgery. However, cell based treatments which intervene and attenuate discopathies and radiculopathies at an earlier time point, providing regenerative and anti-inflammatory treatments could potentially minimize the need of invasive surgery altogether in a select patient population. Research and innovations in the areas of imaging and diagnosis along side of regenerative medicine using stem cells and appropriate biomatrices are essential if progress is to be made in this important field of spinal surgery.

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#### REFERENCES

- [1] NIH. Stem Cell Information. 2008; Available from: <http://stemcells.nih.gov/info/basics/basics2.asp>.
- [2] Thomson JA, Itskovitz-Eldor J, Shapiro SS, *et al.* Embryonic stem cell lines derived from human blastocysts. *Science* 1998; 282(5391): 1145-7.
- [3] Trounson A. Human embryonic stem cells: mother of all cell and tissue types. *Reprod Biomed Online* 2002; 4 Suppl 1: 58-63.
- [4] Rosenfeld J, Bandopadhyay P, Goldschlager T, Brown D. The Ethics of the Treatment of Spinal Cord Injury: Stem Cell Transplants, Motor Neuroprosthetics, and Social Equity. *Top Spinal Cord Inj Rehabil* 2008; 14(1): 76-88.
- [5] Jeffrey S. First Embryonic Stem-Cell-Based Therapy Trial in Spinal-Cord Injury Gets FDA Nod New York: Web MD Professional; 2009; Available from: <http://www.medscape.com/viewarticle/587411>.
- [6] Caplan AI. New Era of Cell-Based Orthopaedic Therapies. *Tissue Eng Part B Rev* 2009.
- [7] MacMillan ML, Davies SM, Nelson GO, *et al.* Twenty years of unrelated donor bone marrow transplantation for pediatric acute leukemia facilitated by the National Marrow Donor Program. *Biol Blood Marrow Transplant* 2008; 14(9 Suppl): 16-22.
- [8] Castro-Malaspina H, Gay RE, Resnick G, *et al.* Characterization of human bone marrow fibroblast colony-forming cells (CFU-F) and their progeny. *Blood* 1980; 56(2): 289-301.
- [9] Friedenstein AJ, Chailakhjan RK, Lalykina KS. The development of fibroblast colonies in monolayer cultures of guinea-pig bone marrow and spleen cells. *Cell Tissue Kinet* 1970; 3(4): 393-403.
- [10] Dennis J, Caplan A. Bone Marrow Mesenchymal Stem Cells. *Stem Cell Handbook*. Albany, New York: Humana Press; 2003. p. 108.
- [11] Zuk PA, Zhu M, Ashjian P, *et al.* Human adipose tissue is a source of multipotent stem cells. *Mol Biol Cell* 2002; 13(12): 4279-95.
- [12] Sakaguchi Y, Sekiya I, Yagishita K, Muneta T. Comparison of human stem cells derived from various mesenchymal tissues: superiority of synovium as a cell source. *Arthritis Rheum* 2005; 52(8): 2521-9.
- [13] Young HE, Steele TA, Bray RA, *et al.* Human reserve pluripotent mesenchymal stem cells are present in the connective tissues of skeletal muscle and dermis derived from fetal, adult, and geriatric donors. *Anat Rec* 2001; 264(1): 51-62.
- [14] Brisby H, Tao H, Ma D, Diwan A. Cell therapy for disc degeneration--potentials and pitfalls. *Orthop Clin North Am* 2004; 35(1): 85-93.
- [15] Wu Y, Chen L, Scott PG, Tredget EE. Mesenchymal stem cells enhance wound healing through differentiation and angiogenesis. *Stem Cells* 2007; 25(10): 2648-59.
- [16] Long X, Olszewski M, Huang W, Kletzel M. Neural cell differentiation *in vitro* from adult human bone marrow mesenchymal stem cells. *Stem Cells Dev* 2005; 14(1): 65-9.
- [17] Dominici M, Le Blanc K, Mueller I, *et al.* Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement. *Cytotherapy* 2006; 8(4): 315-7.
- [18] De Ugarte DA, Alfonso Z, Zuk PA, *et al.* Differential expression of stem cell mobilization-associated molecules on multi-lineage cells from adipose tissue and bone marrow. *Immunol Lett* 2003; 89(2-3): 267-70.
- [19] Locke M, Windsor J, Dunbar PR. Human adipose-derived stem cells: isolation, characterization and applications in surgery. *ANZ J Surg* 2009; 79(4): 235-44.
- [20] Conget PA, Minguell JJ. Phenotypical and functional properties of human bone marrow mesenchymal progenitor cells. *J Cell Physiol* 1999; 181(1):67-73.
- [21] Liu TM, Martina M, Hutmacher DW, *et al.* Identification of common pathways mediating differentiation of bone marrow- and adipose tissue-derived human mesenchymal stem cells into three mesenchymal lineages. *Stem Cells* 2007; 25(3): 750-60.
- [22] Alhadlaq A, Mao JJ. Mesenchymal stem cells: isolation and therapeutics. *Stem Cells Dev* 2004; 13(4): 436-48.
- [23] Ho AD, Wagner W, Franke W. Heterogeneity of mesenchymal stromal cell preparations. *Cytotherapy* 2008; 10(4): 320-30.
- [24] Zannettino AC, Paton S, Arthur A, *et al.* Multipotential human adipose-derived stromal stem cells exhibit a perivascular phenotype *in vitro* and *in vivo*. *J Cell Physiol* 2008; 214(2): 413-21.
- [25] Caplan AI. All MSCs are pericytes? *Cell Stem Cell* 2008; 3(3): 229-30.

- [26] Goldschlager T, Jenkin G, Rosenfeld JV, Ghosh P. Chondrogenic differentiation of adipose-derived stem cells. *ANZ J Surg*. [Letter to the Editor]. 2009; 79(11): 856-7.
- [27] Im GI, Shin YW, Lee KB. Do adipose tissue-derived mesenchymal stem cells have the same osteogenic and chondrogenic potential as bone marrow-derived cells? *Osteoarthritis Cartilage* 2005; 13(10): 845-53.
- [28] Content and Review of Chemistry, Manufacturing, and Control (CMC) Information for Human Somatic Cell Therapy Investigational New Drug Applications, (April 2008).
- [29] Doherty MJ, Ashton BA, Walsh S, *et al.* Vascular pericytes express osteogenic potential *in vitro* and *in vivo*. *J Bone Miner Res* 1998; 13(5): 828-38.
- [30] Risbud MV, Guttapalli A, Tsai TT, *et al.* Evidence for skeletal progenitor cells in the degenerate human intervertebral disc. *Spine* 2007; 32(23): 2537-44.
- [31] Simmons PJ, Gronthos S, Zannettino A, Ohta S, Graves S. Isolation, characterization and functional activity of human marrow stromal progenitors in hemopoiesis. *Prog Clin Biol Res* 1994; 389:271-80.
- [32] Jones P, Gardner L, Menage J, Williams GT, Roberts S. Intervertebral disc cells as competent phagocytes *in vitro*: implications for cell death in disc degeneration. *Arthritis research & therapy* 2008; 10(4): R86.
- [33] Ratajczak MZ, Zuba-Surma EK, Shin DM, Ratajczak J, Kucia M. Very small embryonic-like (VSEL) stem cells in adult organs and their potential role in rejuvenation of tissues and longevity. *Exp Gerontol* 2008; 43(11):1009-17.
- [34] Gronthos S, Graves SE, Ohta S, Simmons PJ. The STRO-1+ fraction of adult human bone marrow contains the osteogenic precursors. *Blood* 1994; 84(12): 4164-73.
- [35] Simmons PJ, Torok-Storb B. Identification of stromal cell precursors in human bone marrow by a novel monoclonal antibody, STRO-1. *Blood* 1991; 78(1): 55-62.
- [36] Gucciardo L, Lories R, Ochsenschein-Kolble N, *et al.* Fetal mesenchymal stem cells: isolation, properties and potential use in perinatology and regenerative medicine. *BJOG* 2009; 116(2): 166-72.
- [37] Siegel N, Rosner M, Hanneder M, Freilinger A, Hengstschlager M. Human amniotic fluid stem cells: a new perspective. *Amino Acids* 2008; 35(2):291-3.
- [38] Serikov V, Hounshell C, Larkin S, *et al.* A BRIEF COMMUNICATION: Human Term Placenta as a Source of Hematopoietic Cells. *Exp Biol Med* (Maywood). 2009; 234(7): 813-23.
- [39] Ilancheran S, Moodley Y, Manuelpillai U. Human fetal membranes: a source of stem cells for tissue regeneration and repair? *Placenta* 2009; 30(1):2-10.
- [40] Lubin BH, Eraklis M, Apicelli G. Umbilical cord blood banking. *Adv Pediatr* 1999; 46: 383-408.
- [41] Ilancheran S, Michalska A, Peh G, *et al.* Stem cells derived from human fetal membranes display multilineage differentiation potential. *Biol Reprod* 2007; 77(3): 577-88.
- [42] Alviano F, Fossati V, Marchionni C, *et al.* Term Amniotic membrane is a high throughput source for multipotent Mesenchymal Stem Cells with the ability to differentiate into endothelial cells *in vitro*. *BMC Dev Biol* 2007; 7:11.
- [43] Soncini M, Vertua E, Gibelli L, *et al.* Isolation and characterization of mesenchymal cells from human fetal membranes. *J Tissue Eng Regen Med* 2007; 1(4): 296-305.
- [44] Armitage JO. Bone marrow transplantation. *N Engl J Med* 1994; 330(12): 827-38.
- [45] Bailo M, Soncini M, Vertua E, *et al.* Engraftment potential of human amnion and chorion cells derived from term placenta. *Transplantation* 2004; 78(10): 1439-48.
- [46] Li H, Niederkorn JY, Neelam S, *et al.* Immunosuppressive factors secreted by human amniotic epithelial cells. *Invest Ophthalmol Vis Sci* 2005; 46(3):900-7.
- [47] Chen HJ, Pires RT, Tseng SC. Amniotic membrane transplantation for severe neurotrophic corneal ulcers. *Br J Ophthalmol* 2000; 84(8): 826-33.
- [48] Pires RT, Chokshi A, Tseng SC. Amniotic membrane transplantation or conjunctival limbal autograft for limbal stem cell deficiency induced by 5-fluorouracil in glaucoma surgeries. *Cornea* 2000; 19(3): 284-7.
- [49] Meller D, Pires RT, Mack RJ, *et al.* Amniotic membrane transplantation for acute chemical or thermal burns. *Ophthalmology* 2000; 107(5): 980-9; discussion 90.
- [50] Goldschlager T, Rosenfeld J, Ghosh P, *et al.* editors. A Comparison Between Allogeneic Mesenchymal Precursor Cells And Amnion Epithelial Cells In Promoting Cervical Intervertebral Fusion In An Ovine Model. Congress of Neurological Surgeons, Annual Meeting; 2009; New Orleans.
- [51] Song L, Tuan RS. Transdifferentiation potential of human mesenchymal stem cells derived from bone marrow. *FASEB J* 2004; 18(9): 980-2.
- [52] Takahashi K, Tanabe K, Ohnuki M, *et al.* Induction of pluripotent stem cells from adult human fibroblasts by defined factors. *Cell* 2007; 131(5): 861-72.
- [53] Yu J, Vodyanik M, Smuga-Otto K, *et al.* Induced Pluripotent Stem Cell Lines Derived from Human Somatic Cells. *Science* 2007; 318(5858): 1917-20.
- [54] Kim D, Kim C, Moon J, *et al.* Generation of Human Induced Pluripotent Stem Cells by Direct Delivery of Reprogramming Proteins. *Cell Stem Cell* 2009; 1-5.
- [55] Maherali N, Hochedlinger K. Guidelines and Techniques for the Generation of Induced Pluripotent Stem Cells. *Cell Stem Cell* 2008; 3(6): 595-605.
- [56] Lian Q, Zhang Y, Zhang J, *et al.* Functional mesenchymal stem cells derived from human induced pluripotent stem cells attenuate limb ischemia in mice. *Circulation* 121(9): 1113-23.
- [57] D'Ippolito G, Diabira S, Howard GA, *et al.* Marrow-isolated adult multilineage inducible (MIAMI) cells, a unique population of postnatal young and old human cells with extensive expansion and differentiation potential. *J Cell Sci* 2004; 117(Pt 14): 2971-81.
- [58] Reyes M, Verfaillie CM. Characterization of multipotent adult progenitor cells, a subpopulation of mesenchymal stem cells. *Ann N Y Acad Sci* 2001; 938: 231-3; discussion 3-5.
- [59] Giordano A, Galderisi U, Marino IR. From the laboratory bench to the patient's bedside: an update on clinical trials with mesenchymal stem cells. *J Cell Physiol* 2007; 211(1): 27-35.
- [60] Kuci S, Kuci Z, Latifi-Pupovci H, *et al.* Adult stem cells as an alternative source of multipotential (pluripotential) cells in regenerative medicine. *Curr Stem Cell Res Ther* 2009; 4(2): 107-17.
- [61] Wojakowski W, Kucia M, Kazmierski M, Ratajczak MZ, Tendera M. Circulating progenitor cells in stable coronary heart disease and acute coronary syndromes: relevant reparatory mechanism? *Heart* 2008; 94(1): 27-33.
- [62] Blattert TR, Delling G, Dalal PS, *et al.* Successful transpedicular lumbar interbody fusion by means of a composite of osteogenic protein-1 (rhBMP-7) and hydroxyapatite carrier: a comparison with autograft and hydroxyapatite in the sheep spine. *Spine* 2002; 27(23): 2697-705.
- [63] Boden SD, Kang J, Sandhu H, Heller JG. Use of recombinant human bone morphogenetic protein-2 to achieve posterolateral lumbar spine fusion in humans: a prospective, randomized clinical pilot trial: 2002 Volvo Award in clinical studies. *Spine* 2002; 27(23): 2662-73.
- [64] Yoon ST, Patel NM. Molecular therapy of the intervertebral disc. *Eur Spine J* 2006; 15 Suppl 3: S379-88.
- [65] Ryu SI, Lim JT, Kim SM, *et al.* Comparison of the biomechanical stability of dense cancellous allograft with tricortical iliac autograft and fibular allograft for cervical interbody fusion. *Eur Spine J* 2006; 15(9): 1339-45.
- [66] Silber JS, Anderson DG, Daffner SD, *et al.* Donor site morbidity after anterior iliac crest bone harvest for single-level anterior cervical discectomy and fusion. *Spine* 2003; 28(2): 134-9.
- [67] Aggarwal S, Pittenger MF. Human mesenchymal stem cells modulate allogeneic immune cell responses. *Blood* 2005; 105(4): 1815.
- [68] Tyndall A, Walker U, Cope A, *et al.* Immunomodulatory properties of mesenchymal stem cells: a review based on an interdisciplinary meeting held at the Kennedy Institute of Rheumatology Division, London, UK, 31 October 2005. *Arthritis research & therapy* 2007; 9(1): 301.
- [69] Ryan JM, Barry FP, Murphy JM, Mahon BP. Mesenchymal stem cells avoid allogeneic rejection. *J Inflamm (Lond)* 2005; 2: 8.
- [70] Maccario R, Podestà M, Moretta A, *et al.* Interaction of human mesenchymal stem cells with cells involved in alloantigen-specific



- immune response favors the differentiation of CD4+ T-cell subsets expressing a regulatory/suppressive phenotype. *Haematologica* 2005; 90(4): 516-25.
- [71] Sonntag VKH, Vollmer DG. Degenerative Disease of the Spine. In: Winn R, editor. *Youmans Neurological Surgery*. New York: Elsevier; 2006 p. 4149-50.
- [72] Mulleman D, Mammou S, Griffoul I, Watier H, Goupille P. Pathophysiology of disk-related sciatica. I.—Evidence supporting a chemical component. *Joint Bone Spine* 2006; 73(2): 151-8.
- [73] Mulleman D, Mammou S, Griffoul I, Watier H, Goupille P. Pathophysiology of disk-related low back pain and sciatica. II. Evidence supporting treatment with TNF-alpha antagonists. *Joint Bone Spine* 2006; 73(3): 270-7.
- [74] Chad DA. Lumbar spinal stenosis. *Neurologic clinics*. 2007; 25(2):407-18.
- [75] Harrop JS, Hanna A, Silva MT, Sharan A. Neurological manifestations of cervical spondylosis: an overview of signs, symptoms, and pathophysiology. *Neurosurgery*. 2007; 60(1 Suppl 1): S14-20.
- [76] Kumar VGR, Madden C, Rea GL. Cervical Spondylotic Myelopathy. In: Winn R, editor. *Youmans Neurological Surgery*. New York: Elsevier; 2006 p. 4447-9.
- [77] Kandel RA, Chang G, Kluge J, Vunjak-Novakovic G, Kaplan D, editors. *How to tissue engineer an intervertebral disc*. World Forum for Spine Research; 2008; Kyoto, Japan: AO Spine International.
- [78] Coppes MH, Marani E, Thomeer RT, Groen GJ. Innervation of "painful" lumbar discs. *Spine*. 1997; 22(20):2342-9; discussion 9-50.
- [79] Peng B, Wu W, Hou S, *et al*. The pathogenesis of discogenic low back pain. *J Bone Joint Surg Br* 2005; 87(1): 62-7.
- [80] Haughton V. Imaging intervertebral disc degeneration. *J Bone Joint Surg Am* 2006; 88 Suppl 2: 15-20.
- [81] Rengachary SS, Balabhadra RS. Black disc disease: a commentary. *Neurosurg Focus* 2002; 13(2): E14.
- [82] Videman T, Battié MC, Gibbons LE, *et al*. Associations between back pain history and lumbar MRI findings. *Spine* 2003; 28(6):582-8.
- [83] Sembrano JN, Polly DW, Jr. How often is low back pain not coming from the back? *Spine* 2009; 34(1): E27-32.
- [84] Witwer BP, Trost GR. Cervical spondylosis: ventral or dorsal surgery. *Neurosurgery* 2007; 60(1 Suppl 1): S130-6.
- [85] Thomé C, Barth M, Scharf J, Schmiedek P. Outcome after lumbar sequestrectomy compared with microdiscectomy: a prospective randomized study. *J Neurosurg Spine* 2005; 2(3): 271-8.
- [86] Carragee EJ, Spinnickie AO, Alamin TF, Paragioudakis S. A prospective controlled study of limited versus subtotal posterior discectomy: short-term outcomes in patients with herniated lumbar intervertebral discs and large posterior anular defect. *Spine* 2006; 31(6): 653-7.
- [87] Jacobs WC, Anderson PG, Limbeek J, Willems PC, Pavlov P. Single or double-level anterior interbody fusion techniques for cervical degenerative disc disease. *Cochrane database of systematic reviews (Online)* 2004; (4): CD004958.
- [88] Irwin ZN, Hilibrand A, Gustavel M, *et al*. Variation in surgical decision making for degenerative spinal disorders. Part II: cervical spine. *Spine* 2005; 30(19): 2214-9.
- [89] Patil PG, Turner DA, Pietrobon R. National trends in surgical procedures for degenerative cervical spine disease: 1990-2000. *Neurosurgery* 2005; 57(4): 753-8; discussion -8.
- [90] Deyo RA. Back surgery—who needs it? *N Engl J Med* 2007; 356(22): 2239-43.
- [91] Resnick DK, Choudhri TF, Dailey AT, *et al*. Guidelines for the performance of fusion procedures for degenerative disease of the lumbar spine. Part 8: lumbar fusion for disc herniation and radiculopathy. *J Neurosurg Spine* 2005; 2(6): 673-8.
- [92] Fritzell P, Hägg O, Wessberg P, Nordwall A, Unknown. 2001 Volvo Award Winner in Clinical Studies: Lumbar fusion versus nonsurgical treatment for chronic low back pain: a multicenter randomized controlled trial from the Swedish Lumbar Spine Study Group. *Spine* 2001; 26(23): 2521-32; discussion 32-4.
- [93] Weatherley CR, Prickett CF, O'Brien JP. Discogenic pain persisting despite solid posterior fusion. *J Bone Joint Surg Br* 1986; 68(1): 142-3.
- [94] Bandopadhyay P, Goldschlager T, Rosenfeld JV. The role of evidence-based medicine in neurosurgery. *J Clin Neurosci* 2008; 15(4): 373-8.
- [95] Dowd GC, Wirth FP. Anterior cervical discectomy: is fusion necessary? *J Neurosurg* 1999; 90(1 Suppl): 8-12.
- [96] Hauerberg J, Kosteljanetz M, Boge-Rasmussen T, *et al*. Anterior cervical discectomy with or without fusion with ray titanium cage: a prospective randomized clinical study. *Spine* 2008; 33(5): 458-64.
- [97] Martins AN. Anterior cervical discectomy with and without interbody bone graft. *J Neurosurg* 1976; 44(3): 290-5.
- [98] Murphy MA, Trimble MB, Piedmonte MR, Kalfas IH. Changes in the cervical foraminal area after anterior discectomy with and without a graft. *Neurosurgery* 1994; 34(1): 93-6.
- [99] Rosenorn J, Hansen E, Rosenorn M. Anterior cervical discectomy with and without fusion. A prospective study. *J Neurosurg*. 1983; 59(2): 252-5.
- [100] van Limbeek J, Jacobs WC, Anderson PG, Pavlov PW. A systematic literature review to identify the best method for a single level anterior cervical interbody fusion. *Eur Spine J* 2000; 9(2): 129-36.
- [101] Bartels RH, Donk R, van der Wilt GJ, Grotenhuis JA, Venderink D. Design of the PROCON trial: a prospective, randomized multicenter study comparing cervical anterior discectomy without fusion, with fusion or with arthroplasty. *BMC Musculoskelet Disord* 2006; 7: 85.
- [102] Urban JP, Smith S, Fairbank JC. Nutrition of the intervertebral disc. *Spine* 2004; 29(23): 2700-9.
- [103] Kandel R, Roberts S, Urban JP. Tissue engineering and the intervertebral disc: the challenges. *Eur Spine J* 2008; 17 Suppl 4: 480-91.
- [104] Floyd T, Ohnmeiss D. A meta-analysis of autograft versus allograft in anterior cervical fusion. *Eur Spine J* 2000; 9(5): 398-403.
- [105] Aho AJ, Hirn M, Aro HT, Heikkilä JT, Meurman O. Bone bank service in Finland. Experience of bacteriologic, serologic and clinical results of the Turku Bone Bank 1972-1995. *Acta Orthop Scand* 1998; 69(6): 559-65.
- [106] Cho DY, Liau WR, Lee WY, *et al*. Preliminary experience using a polyetheretherketone (PEEK) cage in the treatment of cervical disc disease. *Neurosurgery* 2002; 51(6): 1343-49; discussion 9-50.
- [107] Kulkarni AG, Hee HT, Wong HK. Solis cage (PEEK) for anterior cervical fusion: preliminary radiological results with emphasis on fusion and subsidence. *Spine J* 2007; 7(2): 205-9.
- [108] Crawford CH, 3rd, Carreon LY, Djurasovic M, Glassman SD. Lumbar fusion outcomes in patients with rheumatoid arthritis. *Eur Spine J* 2008; 17(6): 822-5.
- [109] Mooney V, McDermott KL, Song J. Effects of smoking and maturation on long-term maintenance of lumbar spinal fusion success. *J Spinal Disord* 1999; 12(5): 380-5.
- [110] Andersen T, Christensen FB, Laursen M, *et al*. Smoking as a predictor of negative outcome in lumbar spinal fusion. *Spine* 2001; 26(23): 2623-8.
- [111] Lumawig JM, Yamazaki A, Watanabe K. Dose-dependent inhibition of diclofenac sodium on posterior lumbar interbody fusion rates. *Spine J* 2009; 9(5): 343-9.
- [112] Schuberth JM, DiDomenico LA, Medicino RW. The utility and effectiveness of bone morphogenetic protein in foot and ankle surgery. *J Foot Ankle Surg* 2009; 48(3): 309-14.
- [113] Smith DM, Cooper GM, Mooney MP, Marra KG, Losee JE. Bone morphogenetic protein 2 therapy for craniofacial surgery. *J Craniofac Surg* 2008; 19(5): 1244-59.
- [114] Rihn JA, Gates C, Glassman SD, *et al*. The use of bone morphogenetic protein in lumbar spine surgery. *J Bone Joint Surg Am* 2008; 90(9): 2014-25.
- [115] Azari K, Doctor JS, Doll BA, Hollinger JO. Bone morphogenetic proteins A review for cranial and maxillofacial surgery. *Oral Maxillofac Surg Clin North Am* 2002; 14(1): 1-14.
- [116] Perri B, Cooper M, Laurysen C, Anand N. Adverse swelling associated with use of rh-BMP-2 in anterior cervical discectomy and fusion: a case study. *Spine J* 2007; 7(2): 235-9.
- [117] Smucker JD, Rhee JM, Singh K, Yoon ST, Heller JG. Increased swelling complications associated with off-label usage of rhBMP-2 in the anterior cervical spine. *Spine* 2006; 31(24): 2813-9.
- [118] FDA. FDA Public Health Notification: Life-threatening Complications Associated with Recombinant Human Bone

- Morphogenetic Protein in Cervical Spine Fusion. Maryland July, 2008; Available from: <http://www.fda.gov/cdrh/safety/070108-rhbm.html>.
- [119] Goldschlager T, Rosenfeld JV, Young IR, Jenkin G. Anterior cervical discectomy and fusion in the ovine model. *J Vis Exp* 2009; 32(1548).
- [120] Goldschlager T, Itescu S, Ghosh P, *et al.* editors. Allogeneic mesenchymal precursor cells safely and effectively increase the rate and robustness of cervical interbody fusion. Orthopedic Research Society, 55th Annual Meeting; 2009; Las Vegas.
- [121] Goldschlager T, Blecher C, Rosenfeld J, *et al.* editors. Quantitative analysis of robust cervical interbody fusion induced by allogeneic mesenchymal precursor cells. Spine Society of Australia, Annual Meeting; 2009; Brisbane.
- [122] Goldschlager T, Itescu S, Ghosh P, *et al.* Cervical Interbody Fusion is Enhanced by Allogeneic Mesenchymal Precursor Cells in an Ovine Model. *Spine* 2010; in press.
- [123] Cheng H, Jiang W, Phillips FM, *et al.* Osteogenic activity of the fourteen types of human bone morphogenetic proteins (BMPs). *J Bone Joint Surg Am* 2003; 85-A(8): 1544-52.
- [124] Goldschlager T. A Phase 1b/2a, Randomized, Double-Blinded, Controlled Study Evaluating Safety and Preliminary Efficacy of NeoFuse™ when Combined with MasterGraft™ Granules in Subjects Undergoing Multi-Level Anterior Cervical Discectomy Model. . Approval number: ACTRN126090010492682009; Available from: <http://www.ANZCTR.org.au/ACTRN12609001049268.aspx>.
- [125] NIH. Study of 3 Doses of NeoFuse Combined With MasterGraft Granules in Subjects Requiring Posterolateral Lumbar Fusion (PLF), NCT00549913. [clinicaltrials.gov](http://clinicaltrials.gov); 2009; Available from: <http://clinicaltrials.gov/ct2/show/NCT00549913>.
- [126] Blumenthal SL, Baker J, Dossett A, Selby DK. The role of anterior lumbar fusion for internal disc disruption. *Spine* 1988; 13(5): 566-9.
- [127] Xie JC, Hurlbert RJ. Discectomy versus discectomy with fusion versus discectomy with fusion and instrumentation: a prospective randomized study. *Neurosurgery* 2007; 61(1): 107-16; discussion 16-7.
- [128] Vaccaro AR, Whang PG, Patel T, *et al.* The safety and efficacy of OP-1 (rhBMP-7) as a replacement for iliac crest autograft for posterolateral lumbar arthrodesis: minimum 4-year follow-up of a pilot study. *Spine J* 2008; 8(3): 457-65.
- [129] Vaccaro AR, Patel T, Fischgrund J, *et al.* A pilot safety and efficacy study of OP-1 putty (rhBMP-7) as an adjunct to iliac crest autograft in posterolateral lumbar fusions. *Eur Spine J* 2003; 12(5): 495-500.
- [130] Nakajima T, Iizuka H, Tsutsumi S, Kayakabe M, Takagishi K. Evaluation of posterolateral spinal fusion using mesenchymal stem cells: differences with or without osteogenic differentiation. *Spine* 2007; 32(22): 2432-6.
- [131] Trials C. Study of 3 Doses of NeoFuse Combined With MasterGraft Granules in Subjects Requiring Posterolateral Lumbar Fusion (PLF), NCT00549913. [clinicaltrials.gov](http://clinicaltrials.gov); 2009; Available from: <http://clinicaltrials.gov/ct2/show/NCT00549913>.
- [132] Koebbe CJ, Maroon JC, Abla A, El-Kadi H, Bost J. Lumbar microdiscectomy: a historical perspective and current technical considerations. *Neurosurg Focus* 2002; 13(2): E3.
- [133] Moore RJ, Vernon-Roberts B, Fraser RD, Osti OL, Schembri M. The origin and fate of herniated lumbar intervertebral disc tissue. *Spine* 1996; 21(18): 2149-55.
- [134] Schaller B. Failed back surgery syndrome: the role of symptomatic segmental single-level instability after lumbar microdiscectomy. *Eur Spine J* 2004; 13(3): 193-8.
- [135] Yorimitsu E, Chiba K, Toyama Y, Hirabayashi K. Long-term outcomes of standard discectomy for lumbar disc herniation: a follow-up study of more than 10 years. *Spine* 2001; 26(6): 652-7.
- [136] McGirt MJ, Ambrossi GL, Dato G, *et al.* Recurrent disc herniation and long-term back pain after primary lumbar discectomy: review of outcomes reported for limited versus aggressive disc removal. *Neurosurgery* 2009; 64(2): 338-44; discussion 44-5.
- [137] Sato M, Asazuma T, Ishihara M, *et al.* An experimental study of the regeneration of the intervertebral disc with an allograft of cultured annulus fibrosus cells using a tissue-engineering method. *Spine* 2003; 28(6): 548-53.
- [138] Hamilton DJ, Seguin CA, Wang J, Pilliar RM, Kandel RA. Formation of a nucleus pulposus-cartilage endplate construct *in vitro*. *Biomaterials* 2006; 27(3): 397-405.
- [139] Gruber HE, Hoelscher GL, Leslie K, Ingram JA, Hanley EN, Jr. Three-dimensional culture of human disc cells within agarose or a collagen sponge: assessment of proteoglycan production. *Biomaterials* 2006; 27(3): 371-6.
- [140] Gruber HE, Leslie K, Ingram J, Norton HJ, Hanley EN. Cell-based tissue engineering for the intervertebral disc: *in vitro* studies of human disc cell gene expression and matrix production within selected cell carriers. *Spine J* 2004; 4(1): 44-55.
- [141] Mizuno H, Roy AK, Vacanti CA, *et al.* Tissue-engineered composites of annulus fibrosus and nucleus pulposus for intervertebral disc replacement. *Spine* 2004; 29(12): 1290-7; discussion 7-8.
- [142] Neidlinger-Wilke C, Wurtz K, Liedert A, *et al.* A three-dimensional collagen matrix as a suitable culture system for the comparison of cyclic strain and hydrostatic pressure effects on intervertebral disc cells. *J Neurosurg Spine* 2005; 2(4): 457-65.
- [143] Desai BJ, Gruber HE, Hanley EN, Jr. The influence of Matrigel or growth factor reduced Matrigel on human intervertebral disc cell growth and proliferation. *Histology and histopathology*. 1999; 14(2): 359-68.
- [144] Gruber HE, Leslie K, Ingram J, *et al.* Colony formation and matrix production by human annulus cells: modulation in three-dimensional culture. *Spine* 2004; 29(13): E267-74.
- [145] Steck E, Bertram H, Abel R, *et al.* Induction of intervertebral disc-like cells from adult mesenchymal stem cells. *Stem cells (Dayton, Ohio)* 2005; 23(3): 403-11.
- [146] Le Visage C, Kim SW, Tateno K, *et al.* Interaction of human mesenchymal stem cells with disc cells: changes in extracellular matrix biosynthesis. *Spine* 2006; 31(18): 2036-42.
- [147] Wei A, Chung SA, Tao H, *et al.* Differentiation of Rodent Bone Marrow Mesenchymal Stem Cells into Intervertebral Disc-Like Cells Following Co-Culture with Rat Disc Tissue. *Tissue Eng Part A* 2009.
- [148] Meisel H, Ganey T, Hutton W, *et al.* Clinical experience in cell-based therapeutics: intervention and outcome. *Eur Spine J* 2006; 15 Suppl 3: S397-405.
- [149] Meisel H, Siodla V, Ganey T, *et al.* Clinical experience in cell-based therapeutics: disc chondrocyte transplantation A treatment for degenerated or damaged intervertebral disc. *Biomol Eng* 2007; 24(1): 5-21.
- [150] Ganey TM, Meisel HJ. A potential role for cell-based therapeutics in the treatment of intervertebral disc herniation. *Eur Spine J* 2002; 11 Suppl 2: S206-14.
- [151] Hohaus C, Ganey TM, Minkus Y, Meisel HJ. Cell transplantation in lumbar spine disc degeneration disease. *Eur Spine J* 2008; 17 Suppl 4: 492-503.
- [152] Ghosh P, Itescu S, Moore R, *et al.* editors. Injection of allogeneic immunoselected Stro-3+ mesenchymal precursor stem into lumbar intervertebral discs attenuates degeneration and promotes the restoration of the disc extracellular matrix. An experimental study in an ovine model of disc degeneration. *Osteoarthritis Research Society International*; 2009; Montreal.
- [153] Hilibrand AS, Carlson GD, Palumbo MA, Jones PK, Bohlman HH. Radiculopathy and myelopathy at segments adjacent to the site of a previous anterior cervical arthrodesis. *J Bone Joint Surg Am* 1999; 81(4): 519-28.
- [154] Auerbach JD, Jones KJ, Frasca CI, *et al.* The prevalence of indications and contraindications to cervical total disc replacement. *Spine J*. 2007.
- [155] Bertagnoli R, Yue JJ, Pfeiffer F, *et al.* Early results after ProDisc-C cervical disc replacement. *J Neurosurg Spine* 2005; 2(4): 403-10.
- [156] Coric D, Finger F, Boltes P. Prospective randomized controlled study of the Bryan Cervical Disc: early clinical results from a single investigational site. *J Neurosurg Spine* 2006; 4(1): 31-5.
- [157] Hacker R. Cervical disc arthroplasty: a controlled randomized prospective study with intermediate follow-up results. Invited submission from the joint section meeting on disorders of the spine and peripheral nerves. *J Neurosurg Spine* 2005; 3(6): 424-8.
- [158] Porchet F, Metcalf N. Clinical outcomes with the Prestige II cervical disc: preliminary results from a prospective randomized clinical trial. *Neurosurg Focus* 2004; 17(3): E6.

- [159] Ruan D, He Q, Ding Y, *et al.* Intervertebral disc transplantation in the treatment of degenerative spine disease: a preliminary study. *Lancet* 2007; 369(9566): 993-9.
- [160] Goldschlager T, Ghosh P, Wu J, *et al.* Regeneration of the cervical intervertebral disc Part 1: Enhanced proliferation and chondrogenic differentiation of Mesenchymal Precursor Stem Cells (MPC) cultured in collagen sponges in the presence of Pentosan Polysulfate (PPS). Submitted 2009.
- [161] Goldschlager T, Ghosh P, Zannettino A, *et al.* Cervical Motion Preservation using Mesenchymal Progenitor Cells and a novel chondrogenic agent, Pentosan Polysulfate - A preliminary study in an ovine model. *Neurosurgical Focus* 2010 (in press).
- [162] Abbed KM, Coumans JV. Cervical radiculopathy: pathophysiology, presentation, and clinical evaluation. *Neurosurgery* 2007; 60(1 Suppl 1): S28-34.
- [163] Melrose J, Roberts S, Smith S, Menage J, Ghosh P. Increased nerve and blood vessel ingrowth associated with proteoglycan depletion in an ovine anular lesion model of experimental disc degeneration. *Spine* 2002; 27(12): 1278-85.
- [164] Finch P. Technology Insight: imaging of low back pain. *Nat Clin Pract Rheumatol* 2006; 2(10): 554-61.
- [165] Derby R, Kim BJ, Lee SH, *et al.* Comparison of discographic findings in asymptomatic subject discs and the negative discs of chronic LBP patients: can discography distinguish asymptomatic discs among morphologically abnormal discs? *Spine J* 2005; 5(4): 389-94.
- [166] Buenaventura RM, Shah RV, Patel V, Benyamin R, Singh V. Systematic review of discography as a diagnostic test for spinal pain: an update. *Pain physician* 2007; 10(1): 147-64.
- [167] Jensen MC, Brant-Zawadzki MN, Obuchowski N, *et al.* Magnetic resonance imaging of the lumbar spine in people without back pain. *N Engl J Med* 1994; 331(2): 69-73.
- [168] Freemont AJ, Peacock TE, Goupille P, *et al.* Nerve ingrowth into diseased intervertebral disc in chronic back pain. *Lancet* 1997; 350(9072): 178-81.

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